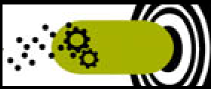

BACTERIAL CROWDING – UPO Sevilla

QUANTITATIVE	CAPILLARY ASSAY USING FLOW-CHAMBERS

1. ***E. Coli*** **inocula in triptone broth in low shaking (between 100-200 rpm/min)**

   Inocula must be incubated at low temperature and low shaking in order to achieve a peak development of flagella. High shaking might provoke the loss of flagella; the production of flagella wouldn't be possible in rich environments since bacteria wouldn't need them.

2. **Taking a sample from one inoculum and looking at the motility under the microscope.**

   It is crucial to observe that flagella have been properly developed in the bacteria; in the same way it is also important to see that those bacteria are mobiles. If the final result is a low motility, the assay should be restarted.

3. **Diluting the** ***E. Coli*** **culture 100 times.**

   We dilute the triptone broth in E. Coli culture a hundred of times. Incubate again at 30ºC in low shaking till exponential medium phase. The final volume would be 20 ml so we will have to inoculate 200 μl.

4. **Check motility under the microscope. Again, we check the motility to observe that it still exits.**

5. **Wash twice the culture centrifugating and resuspending in chemotaxis buffer.**

   It's essential to be really careful when resuspending pellets, it is needed to hit softly in the base of the tube. The centrifuges must be done in a low speed to prevent the loss of flagella (10 min in 8000 G)

6. **Measure optical density and adjust the volume in order to achieve around 10^7 cfu/ml (optical density ~0,01).**

7. **Assay set-up**

   - *Sterilize two flow chambers using UV.*
   - *Fill 7 capillaries with chemotaxis buffer and other 7 with the chemoattractant.*
   - *Fill the flow-chamber channel with 100µl of cell suspension. Put two capillaries in each channel, one with chemoattractant and the other with chemotaxis buffer.*
   - *Spread $2x10^3$, $2x10^4$ and $2x10^5$ dilutions over LB+Ap plates.*

- *Put 200µl of suspension into the Adler chamber. Place capillaries over it and look at under the microscope at t=0,15,20,45 y 60.*

8. **Incubation at 30ºC for 1h.**

9. **Dilution and spread in LB plates.**

Once the capillaries have been taken away from the gel, the dilution and spread in LB plates is started capillaries (we use 2 10^3, 2 10^4 and 2 10^5 dilution factor). The capillary content is obtained in a 1.5mL tube by centrifugation: The capillary content is obtained in a 1.5mL tube by centrifugation: Wash the outside of capillaries using distilled water and then break the seal and empty capillaries into 1.5mL tubes that contain 200 µl of chemotaxis buffer; after that , centrifuge it. LB plates are incubated overnight.

IMPORTANT: To work with *Pseudomonas* it is necessary to set up inocula in minimal medium instead of triptone broth. Plus, the culture must rest till it achieves the exponential late phase, since it is in here when flagella are developed. At the end of this assay the final incubation must be done at 30ºC.