7.2.10
Meeting Minutes
Meeting with Advisors

Don't expose our gels so much, keep intensity a bit lower.

Do gel extractions, grab the insert from the gel and use that to ligate so we don't have the binding competition from the plasmid that originally contained the insert. Zymo gel extraction kit works best. But we can grab qiagen as well. Want about 4 times as much insert as backbone, at least 1:1 insert:backbone. We can use the kit from the south side. Try to use bigger combs, low melt agarose, be careful about taking the comb out to not tear the gel, stick it in the fridge for a little while to make it easier to pull it out without tearing.

Maybe contaminated primers with template. Be really careful about contaminating things, change tips is anything touches it. Try just running primers and see what we get. Test the pipette by just sucking up and pipetting out, run it. We'll see about getting a guy out here to calibrate them.

For synthesizing promoter/RBS: use oligos, 2 rounds of primer extension, purify on acrylamide gel. Design 250 bp primer and use that to synthesize the promoter/RBS.

60 base pairs is pretty much the max we can use without purifying for primers.

cut our plasmids before running them on the gel to be able to compare them the ladders since everything is standardized against linear DNA.

We need to start asking companies for next year's funding around this time.

GAMES camp—meeting on July 19th in the morning. Want more images, more slides so one concept per slide.

RpoS promoter for the RpoS gene to use as stationary phase promoter for collection system.

We can do a membrane prep to see if the protein fusions are in the membrane.

Dr. Jin has a kit for the protein gels, we can buy pre-casted gels. We can talk to Lon? About outer membrane protein gels.

We can talk to microanalytic people in chemistry about metal assays. Check on school of chem's website.