

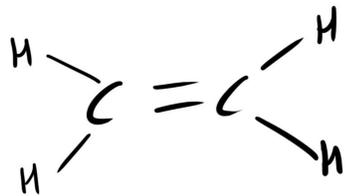
Design and Construction of an Ethylene Generator

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Introduction

We rely heavily on plastics in our day to day lives – and yet they are currently produced from a non-renewable resource, fossil fuels, in a process requiring large amount of energy. We have found that the majority of plastic products are produced from or derived from the compound Ethylene.

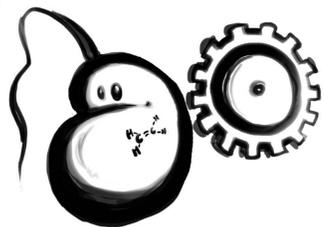


The Ethylene molecule



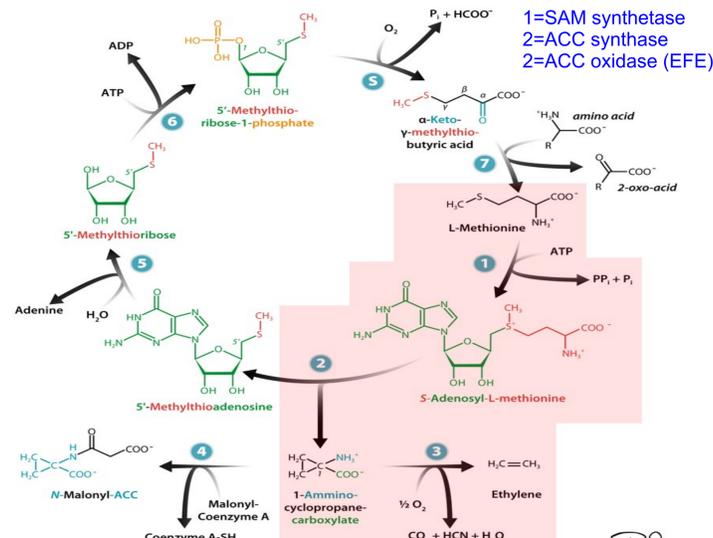
A steam cracking tower

The problem: Ethylene is currently produced in an energy intensive process called steam cracking where large hydrocarbons from oil or natural gas are heated to roughly 900 °C and saturated with steam and subsequently cooled to -157 °C to be compressed and distilled multiple times. This process has a large carbon footprint. To solve this, we have found that plants can produce ethylene from the amino acid methionine at room temperature with renewable and readily available compounds at a much lower carbon cost.



Aim: Construct an ethylene generation device in an *Escherichia coli* chassis to produce ethylene using the enzymes involved in plant ethylene biosynthesis.

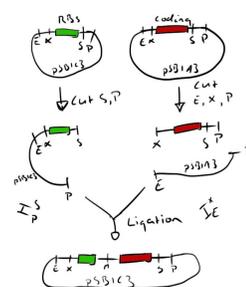
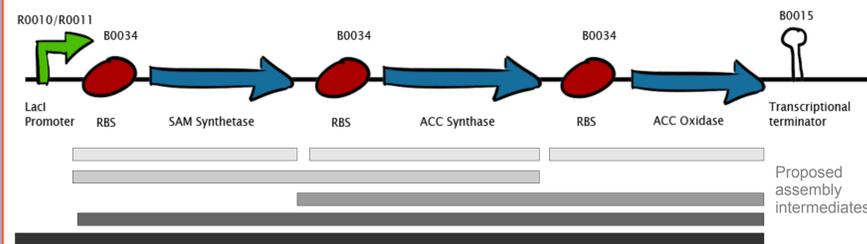
Ethylene biosynthesis



Modified from: <http://commons.wikimedia.org/wiki/File:Yang-cycle.png> (CC-SA 3.0)

Ethylene biosynthesis occurs in plants through the Yang cycle, also known as the methionine cycle.

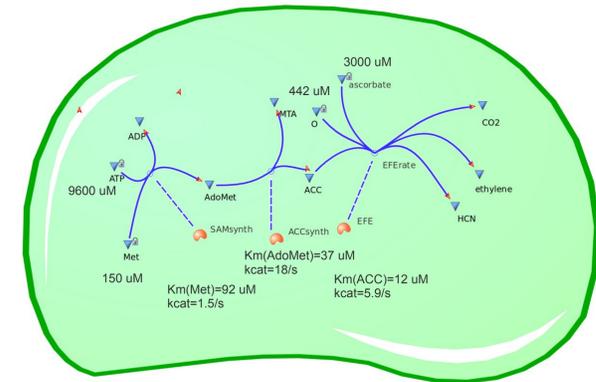
Design & Assembly



Alternative RBS assembly method

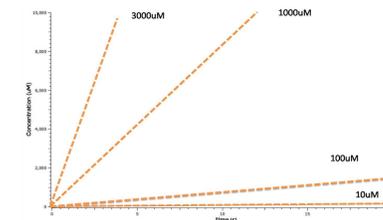
- Design uses Lac inducible promoter (R0011) and strong ribosome binding sites (RBS) (B0034)
- Initially tried 3A Assembly procedure
- Experienced problems with attaching RBS to coding sequences
 - RBS sequence too small?
 - RBS lost?
- Developed alternative assembly procedure (See left) to try and compensate for the possibility of the RBS being lost
- Successfully cloned SAM Synthetase from *E. coli* genomic DNA
 - Shipped as part K417000

Kinetic Modelling



Used the program Tinkercell to create a simple model of the device.

Estimated ethylene output



Used Michaelis Menton Kinetics to model the rate of reaction

Fixed input and enzyme concentrations

Future Work

- Finish construction of our design
- Measure ethylene output via Gas Chromatography
- HCN removal via the enzyme Cyanide Dihydratase
- Reconstruct entire Yang's Cycle to recycle methionine
- Optimise ethylene output via alternative RBSs, promoters
- Measure enzyme concentration, steady state



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